

# Legionella BCYE Agar Base

Art. No. 01-687

## Also known as

CYE

## Specification

Solid medium base used for the detection, isolation and enumeration of *Legionella* from water according to the ISO standards 11731:1998 and 11731-2:2004.

## Formula\* in g/L

Activated Charcoal.....	2,00
Yeast Extract.....	10,00
Agar.....	15,00
Final pH 6,9 ± 0,2 at 25°C	

\* Adjusted and /or supplemented as required to meet performance criteria

## Directions

Suspend 13,5 g of powder in 500 mL of distilled water and bring to the boil dissolving completely. Sterilize by autoclaving at 121°C for 15 minutes. Allow to cool to 47-50°C and add aseptically a reconstituted vial (Art. No. 06-137-LYO) of *Legionella* BCYE Growth Supplement. Mix gently and pour into Petri dishes. If a selective medium for *Legionella* is desired, it can be obtained by the addition of a vial (Art. No. 06-138-LYO) of *Legionella* GVPC Selective Supplement to 500 mL of BCYE Medium melted and cooled to 47-50°C.

If the control medium BCYE – Cys is desired it can be obtained by the addition of a reconstituted vial of (Art. No. 06-134-LYO) *Legionella* BCYE w/o Cysteine Growth Supplement to the sterile, melted and cooled *Legionella* BCYE Agar Base.

## Description

The actual formulation of this medium is according to the ISO Standards 11731 and 11731-2, but BCYE Agar is based in a modification of a previously described media. In 1979 Feeley and collaborators described Charcoal Yeast Extract (CYE) Agar as a modification of the F-G Agar. They replaced the starch in the F-G Agar with activated charcoal and substituted yeast extract for casein hydrolysate, resulting in a better recovery of *Legionella pneumophila*. Pasculle, in 1980, reported that CYE Agar could be improved by buffering the medium with ACES buffer and a year later Edelstein increased the sensitivity of the medium by adding α-ketoglutarate which is the present formulation (BCYE Agar).

The medium consist of a Medium base supplemented with growth factors (BCYE Agar) and the Selective Medium supplemented with inhibitors of undesirable accompanying flora. The yeast Extract supplies the basic nutrients as the medium contains no fermentable carbohydrates. L-Cysteine, Ferric pyrophosphate and α-ketoglutarate are incorporate to satisfy the specific nutritional requirements of *Legionella* species.

The activated charcoal decomposes hydrogen peroxide, a toxic metabolic product, and may also collect CO<sub>2</sub> and modify surface tension. The addition of the buffer helps maintain the proper pH for optimal growth. The selectivity is increased by the addition of Vancomycin and polymyxin B which inhibit Gram-positive bacteria and cycloheximide or natamycin which are antifungal agents and inhibits the yeast growth.

## Technique

Refer to the ISO Standards 11731 and 11731-2 or other standard procedures to obtain isolated colonies from specimens and samples.

Allow the inoculated plates to stand until the inocula has been absorbed. Invert the plates and incubate at 36 ± 1°C for up to 10 days. To ensure the atmosphere in the incubator is humid, place a tray of water in the bottom of the incubator. Top up this tray with fresh water (if necessary) each time the plates are examined. Incubation in an atmosphere of air with 2,5% (volume fraction) CO<sub>2</sub> may be beneficial for the growth of some *Legionella*, but it is not essential.

Examine the plates with a plate microscope on at least three occasions at intervals of 2 to 4 days during the 10-day incubation period, as *Legionella* grow slowly an can be masked by the growth of other organisms. Record the number of each type of colony present.

Colonies of *Legionella* are often white-grey-blue-purple in colour, but may be brown, pink, lime-green or deep-red. They are smooth with a smooth edges and exhibit a characteristic ground-glass appearance. Under ultraviolet light colonies of several species autofluoresce brilliant white, but others are red and *L. pneumophila* appear dull green often tinged with yellow. All presumptive colonies must be confirmed by cultural, biochemical, serological or genetic methods.

## Necessary supplements

### *Legionella* BCYE Growth Supplement (Art. No. 06-137-LYO)

ACES Buffer.....	3,600 g
Potassium hydroxide.....	1,400 g
Ferric pyrophosphate.....	0,125 g
L-Cysteine HCl.....	0,200 g
Potassium α-ketoglutarate.....	0,500 g
Distilled water (Solvent)	

### *Legionella* GVPC Selective Supplement (Art. No. 06-138-LYO) and

Vancomycin.....	0,50 mg
Polymyxin B sulfate.....	40000,00 IU
Cycloheximide.....	40,00 mg
Glycine (ammonia free).....	1,50 g
Distilled water (Solvent)	

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# Legionella BCYE Agar Base

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## Legionella BCYE w/o Cysteine NO Growth Supplement (Art. No. 06-134-LYO)

Vial Contents:

Necessary amount for 500 mL of complete medium.

ACES Buffer.....	3,600 g
Potassium hydroxide.....	1,400 g
Ferric pyrophosphate.....	0,125 g
Potassium $\alpha$ -ketoglutarate.....	0,500 g
Distilled water (Solvent)	

## References

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- ISO 11731 Standard (1998) Water Quality – Detection and Enumeration of *Legionella*.

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- ISO/TS 11133-1: 2009. Microbiology of food and animal feeding stuffs.- Guidelines on preparation and production of culture media. Part 1: General guidelines on quality assurance for the preparation of culture media in the laboratory.
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- WARD, K.W. (1995) Processing and interpretation of specimens for *Legionella* spp. In "Clinical Microbiology Procedures Handbook" Chap. 12.1 edited b H.D. Isenberg. ASM Press. Washington DC, USA.

## Storage

For laboratory use only. Keep tightly closed, away from bright light, in a cool dry place (+4°C to 30°C and <60% RH).

## Quality control

**Incubation temperature:** 35°C  $\pm$  2,0

**Incubation time:** 3 - 10 days

**Inoculum:** 150-300 CFU (Productivity) // 1.000-10.000 CFU (Selectivity). Spiral Plate Method (ISO 11133-1/2)

Microorganism	Growth	Remarks
<i>Legionella pneumophila</i> ATCC 33152	Good	Grey - white colonies
<i>Staphylococcus epidermidis</i> ATCC 12228	Inhibited	-