# **Bacillus cereus Selective Agar**

#### Art. No. 01-487

#### Also known as

PEMBA (Polymyxin Egg yolk Manitol Blue Agar)

#### Specification

Selective solid medium for the enumeration of *Bacillus cereus* in food, according to ISO 21871 and NMKL 674 standards.

#### Formula\* in g/L

Peptone	
Mannitol	10,00
Sodium chloride	2,00
Magnesium sulfate	0,20
Disodium phosphate	2,50
Potassium phosphate	0,25
Brom thymol blue	0,12
Sodium pyruvate	10,00
Agar	14,00
Final pH 7.2 ± 0.2 at 25°C	

Final pH 7,2  $\pm$  0,2 at 25°C

#### **Directions**

Suspend 40 g of powder in 950 mL of distilled water. Allow it to soak and bring to the boil. Distribute into suitable containers and sterilize in the autoclave at 121°C for 15 minutes. Let it cool to 50°C and then add 50 mL/L of Egg Yolk Sterile Emulsion (Art. No. 06-016) and 2 vials/Litre of Polymyxin B Sulfate (Art. No. 06-021CASE or 06-021-LYO) to obtein a 100 U/mL concentration. Homogenize and pour into plates.

#### Description

Bacillus cereus Selective Agar is formulated according to the Food Analysis Nordic Committee (NMLK) standards. This standard uses this medium and Blood Agar Base (Art. No. 01-352) simultaneously for the detection and enumeration of *B. cereus* in any type of food. This medium can also be used to confirm presumptive colonies, in this instance Polymyxin may be omitted.

#### **Technique**

NMLK proposes the simultaneous use of *Bacillus cereus* Selective Agar and Blood Agar Base (Art. No. 01-352). Both media are inoculated by surface streaking with 0,1 mL aliquots which are spread with a Drigalsky loop. Both series of plates are incubated at 30°C for 24 hours.

Typical *B. cereus* colonies in Blood Agar are big, irregular, dirty-white or grey with a surrounding halo of haemolysis. With *B.cereus* Selective Agar, colonies are blue, surrounded by a clear zone of egg yolk digestion (lecithinase positive).

If there is an equal amount of typical colonies on both the media, confirmative tests may not be necessary.

#### **Necessary supplements**

Polymyxin B Sulfate Selective Supplement (Art. No. 06-021CASE / 06-021-LYO)

Vial Contents:

Polymyxin B sulfate	50000,00 I	U
Mannitol (excipient)	100,00 m	ıg

Distilled water (Solvent)

#### References

- CORRY, J.E.L., G.D.W. CURTIS & R.M. BAIRD (2003) Handbook of Culture Media for Food Microbiology. Elsevier Sci. B.V. Amsterdam. The Netherlands.
- FIL-IDF 181:1998 Provisional Int. Standard. Dried Milk Products.
  Enumeration of Bacillus cereus. Most probable number technique.
- ISO/TS 11133-1: 2009. Microbiology of food and animal feeding stuffs.-Guidelines on preparation and production of culture media. Part 1: General guidelines on quality assurance for the preparation of culture media in the laboratory.
- ISO/TS 11133-2: 2003 Corr. 2004. Microbiology of food and animal feeding stuffs.- Guidelines on preparation and production of culture media. Part 2: Practical guidelines on performance testing of culture media.
- ISO 21871 Standard (2006) Microbiology of food and feeding stuffs.-Horizontal method for the determination of low numbers of presumptive Bacillus cereus.- Most probable number technique and detection method.
- NORDISK METODIK KOMITE FÖR LIVSMEDEL (1997) UDC 570.852.11 #674ntq.

#### Storage

For laboratory use only. Keep tightly closed, away from bright light, in a cool dry place (+4°C to 30°C and <60% RH).

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<sup>\*</sup> Adjusted and /or supplemented as required to meet performance criteria

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### **Quality control**

Incubation temperature: 30°C ± 2,0

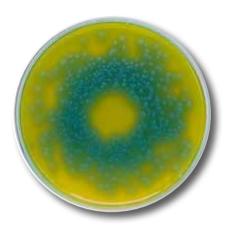
Incubation time: 24 - 48 h

Inoculum: 10-100 CFU (Productivity) // 1.000-10.000 CFU (Selectivity). Spiral Plate Method (ISO/TS 11133-1/2)

Microorganism	Growth	Remarks
Bacillus subtilis ATCC 6633	Fair to good	-
Bacillus cereus var. mycoides ATCC 11778	Productivity > 0.70	White colonies w. precipitate. Green Medium
Bacillus cereus ATCC 10876	Productivity > 0.70	White colonies w. precipitate. Green Medium
Pseudomonas aeruginosa ATCC 27853	Inhibited	<u>.</u>



Uninoculated Plate (Control)



Bacillus cereus var. mycoides ATCC 11778